Enzymatic Ring-Opening Polymerization of Lactones Catalyzed by Lipase

## Hiroshi UYAMA and Shiro KOBAYASHI\*

Department of Molecular Chemistry and Engineering, Faculty of Engineering, Tohoku University, Aoba, Sendai 980

Enzymatic ring-opening polymerization of lactones was achieved by using lipase as catalyst. The polymerization of ε-caprolactone by *Pseudomonas fluorescens* lipase at 60 °C in bulk for 10 days afforded a polyester with average molecular weight of 7.0x10<sup>3</sup>. From <sup>1</sup>H and <sup>13</sup>C NMR analysis, the polymer possesses the terminal structure of a carboxylic acid group at one end and a hydroxyl group at the other.

Organic reactions by enzyme catalysts have been extensively studied because of high substrate specificity and enantioselectivity. By utilizing such specific properties of enzymes, a polymerization catalyzed by enzymes (enzymatic polymerization) has much potential for synthesis of new functional polymers. However, there have been a few reports on enzymatic polymerizations, and moreover, the enzymatic catalysis has not been utilized fully in the polymerization chemistry. Recently, cellulose was successfully synthesized for the first time via a non-biosynthetic path by enzymatic polymerization of  $\beta$ -D-cellobiosyl fluoride monomer using cellulase as catalyst. Further, we reported a new type of enzymatic polymerization; an oxidation polymerization of  $\alpha$ -phenylenediamine catalyzed by horseradish peroxidase produced a polymer consisting of iminophenylene unit, which is unable to be obtained by a conventional oxidation polymerization. Almost all the enzymatic polymerizations hitherto reported were of polycondensation type. In this study, we have broadened the scope of enzymatic polymerization to ring-opening polymerization of lactones by lipase catalyst.

The polymerization of lactones was carried out in bulk for 10 days. In this study,  $\varepsilon$ -caprolactone ( $\varepsilon$ -CL) and  $\delta$ -valerolactone ( $\delta$ -VL) were employed as a monomer and lipase enzymes used were derived from *Pseudomonas fluorescens* (lipase P), *Candida cylindracea* (lipase B), and porcine pancreas (PPL). Polymerizations gave a corresponding ring-opened type polyester from  $\varepsilon$ -CL and  $\delta$ -VL, respectively (Table 1). All the enzymes examined were effective for the polymerization of  $\varepsilon$ -CL. Among them, lipase P afforded the

polyester of higher molecular weight in a better yield.

The effect of the temperature on the polymerization of  $\varepsilon$ -CL was examined (Entries 1 and 4-6). The higher the polymerization temperature, the higher the monomer conversion and the molecular weight of the polymer. Lipase P was also effective for the polymerization of  $\delta$ -VL (Entries 7 and 8). The rate of the monomer conversion was larger than that of  $\varepsilon$ -CL, however the molecular weight of the polymer was lower.

Table 1. Enzymatic polymerzation of lactones catalyzed by lipase<sup>a)</sup>

Entry	Lactone	Enzyme	Temp (°C)	Conv.	$M_{\rm n}^{\rm b)}$ (x10 <sup>-3</sup> )	$M_{\rm w}/M_{\rm n}^{\rm b)}$
1	ε-CL	lipase P	60	85	7.0	2.2
2	ε-CL	lipase B	60	75	3.3	2.5
3	ε-CL	PPL	60	69	2.5	1.9
4	ε-CL	lipase P	30	8	1.1	1.4
5	ε-CL	lipase P	45	40	3.4	2.5
6	ε-CL	lipase P	75	92	7.7	2.4
7	δ-VL	lipase P	45	95	1.6	2.4
8	δ-VL	lipase P	60	95	1.9	3.0

a) Polymerization of lactone (1 mmol) by lipase (50 mg) in bulk for 10 days. b) Determined by GPC.

In order to examine the structure of terminal groups,  $^{13}$ C and  $^{1}$ H NMR analysis of the polymer (Entry 1) was performed.  $^{13}$ C NMR spectrum of poly( $\varepsilon$ -CL) shows three small characteristic peaks besides main peaks due to the carbons of poly( $\varepsilon$ -CL); peaks at  $\delta$  177, 32, and 62 ascribable to the carbon of carboxylic acid group, the carbon adjacent to the carboxylic acid group, and the  $\alpha$ -carbon of hydroxyl group, respectively. In the  $^{1}$ H NMR spectrum, a small triplet peak at  $\delta$  3.5 due to the  $\alpha$ -methylene protons of the hydroxyl group is observed in addition to the main peaks of poly( $\varepsilon$ -CL). From the integrated area of the peaks due to the  $\alpha$ -methylene protons of the hydroxyl group and to C(=0)OCH<sub>2</sub>C ( $\delta$  4.0) of the polymer main-chain, the molecular weight of the polymer was calculated as  $7.1 \times 10^{3}$ , which is close to that determined by GPC. These data support the terminal structure of the polymer having a carboxylic acid group at one end and a hydroxyl group at the other.

Further investigations including the mechanism of the present polymerization are now in progress.

We thank Mr. K. Takeya in our laboratory for experimental assistance.

## References

- 1) J. B. Jones, Tetrahedron, 42, 3351(1986).
- 2) S. Kobayashi, S. Shoda, and H. Uyama, Adv. Polym. Sci., in press.
- 3) S. Kobayashi, K. Kashiwa, T. Kawasaki, and S. Shoda, J. Am. Chem. Soc., 113, 3079(1991).
- 4) S. Kobayashi, I. Kaneko, and H. Uyama, Chem. Lett., 1992, 393.
- 5) This study has been orally presented: S. Kobayashi and H. Uyama, The 65th Annual Meeting, Chem. Soc. Jpn. Prepr., vol. II, 270(1993).

(Received April 15, 1993)